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The Adhesive Protein cDNA of *Mytilus galloprovincialis* Encodes Decapeptide Repeats but No Hexapeptide Motif

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Abstract. A mussel is attached to hard surfaces by its byssus, which consists of a bundle of threads, each with a fibrous collagenous core coated with adhesive proteins. We constructed a cDNA library from RNA isolated from the foot of the mussel *Mytilus galloprovincialis* sampled in Japan. The library was probed with a nucleotide sequence corresponding to a part of the decapeptide repeat motif in the major adhesive protein of the closely related species *M. edulis*, and a clone including the whole coding region of the same adhesive protein of *M. galloprovincialis* was isolated. The sequences of the signal and nonrepetitive regions of the protein of *M. galloprovincialis* were homologous to those of *M. edulis*, despite several substitutions and a deletion of 18 amino acids. The repetitive region included a tetradecapeptide sequence and 62 repeats of the same decapeptide motif as in *M. edulis*, but hexapeptide sequences present in *M. edulis* were absent in the protein of *M. galloprovincialis*. In the decapeptide motif, two tyrosine residues, two lysine residues, and one of the two proline residues were highly conserved, but other residues were frequently substituted. In some residues in the decapeptide motif, specific codon usages were observed, suggesting that the nucleotide sequence itself has a function.

Introduction

Mussels in the genus *Mytilus* are distributed globally in temperate marine intertidal zones. They attach themselves to solid intertidal surfaces by means of the byssus. The byssus is a bundle of threads each consisting mainly of a fibrous collagenous core coated by adhesive proteins. The protein components of the byssus have been extensively studied in *M. edulis* (Waite, 1987, 1992, for reviews). A major adhesive protein is a 130 kDa protein containing a high proportion of 3,4-dihydroxyphenylalanine (DOPA) residues (Waite and Tanzer, 1981; Waite, 1983). The protein is reported to be largely composed of tandem repeats of the decapeptide Ala-Lys-Pro-Ser-Tyr-Hyp-Hyp-Thr-DOPA-Lys, where Hyp is 3- or 4-hydroxyproline (Waite *et al.*, 1985). Other mussel species have similar proteins, each with a unique repeat motif (Waite, 1986; Waite *et al.*, 1989; Rzepecki *et al.*, 1991). Partial sequences of cDNA and genomic DNA encoding the adhesive protein were reported in *M. edulis* (Strausberg *et al.*, 1989; Filpula *et al.*, 1990). The complete amino acid sequence of the adhesive protein from *M. edulis* has been deduced from its cDNA (Laursen, 1992). These studies showed that this adhesive protein contains more than 80 tandem repeats, of which more than 70 are decapeptides and others are hexapeptides.

In this study, we isolated a cDNA clone containing the whole coding region of the adhesive protein from another major species of mussel, *M. galloprovincialis*, which is closely related to *M. edulis* (Gosling, 1984; Gardner, 1992; Geller *et al.*, 1993). We have found that the cDNA encodes a polypeptide containing 62 repeats of the decapeptide found in *M. edulis*, as well as a tetradecapeptide, but no hexapeptide repeat.

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Materials and Methods

Isolation of mRNA

Mussels (*M. galloprovincialis*) about 4 cm in shell length were sampled at Miyako Bay, Iwate prefecture, Japan. The foot was isolated from 12 mussels and the total RNA

Figure 1. Northern blot analysis of RNA extracted from the foot of *Mytilis galloprovincialis*. One microgram of RNA was electrophoresed on a 1% agarose gel, transferred onto a nylon membrane, and hybridized with the oligonucleotide probe corresponding to a part of the decapeptide sequence. Allowheads indicate the position of 18S and 28S rRNA.

was extracted using the Total RNA Separator Kit (Clontech Laboratories, Palo Alto, CA). Poly(A)$^+$RNA was isolated using the mRNA separator (Clontech Laboratories, Palo Alto, CA).

Northern blot hybridization

Poly(A)$^+$RNA was electrophoresed on a 1% agarose gel, transferred onto a nylon membrane, and hybridized with a [^{32}P]ATP-labeled oligonucleotide probe,

ATA(T,A)GTTGGAGGATAA(C,G)TTGGCTT,

that corresponds to a part of the antisense sequence of the decapeptide repeats of *M. edulis* (Strausberg *et al.*, 1989).

Screening of the cDNA library

cDNA was synthesized using the cDNA Synthesis Kit Plus (Amersham). A cDNA library was constructed using the cDNA cloning system lambda gt10 (Amersham). The library was screened using the same probe used for the northern blotting. Ten positive clones were picked up and the size of inserts was determined by excising with *Eco*RI. The longest insert was subcloned into a plasmid vector, BluescriptIISK+ (Stratagene). Restriction analysis of the BluescriptIISK+ subclone was performed using *Apa*I, *Bam*HI, *Eco*RI, *Hinc*II, *Hind*III, *Kpn*I, *Not* I, *Pvu*II, *Pst*I, *Sac*I, *Sal*I, *Sca*I, *Sma*I, *Spe*I, *Xba*I and *Xho*I.

Sequencing

To determine the whole sequences of both strands of the insert, the plasmid containing the insert was digested with *Apa*I/*Hin*dIII or *Sac*I/*Xba*I, and deletion derivatives were produced using the Kilo-Deletion Sequence Kit (Takara, Kyoto, Japan). The original subclone, 28 *Apa*I/*Hin*dIII-generated clones, and 17 *Sac*I/*Xba*I-generated clones were sequenced using a 373A DNA sequencer (Applied Biosystems Inc.).

Results

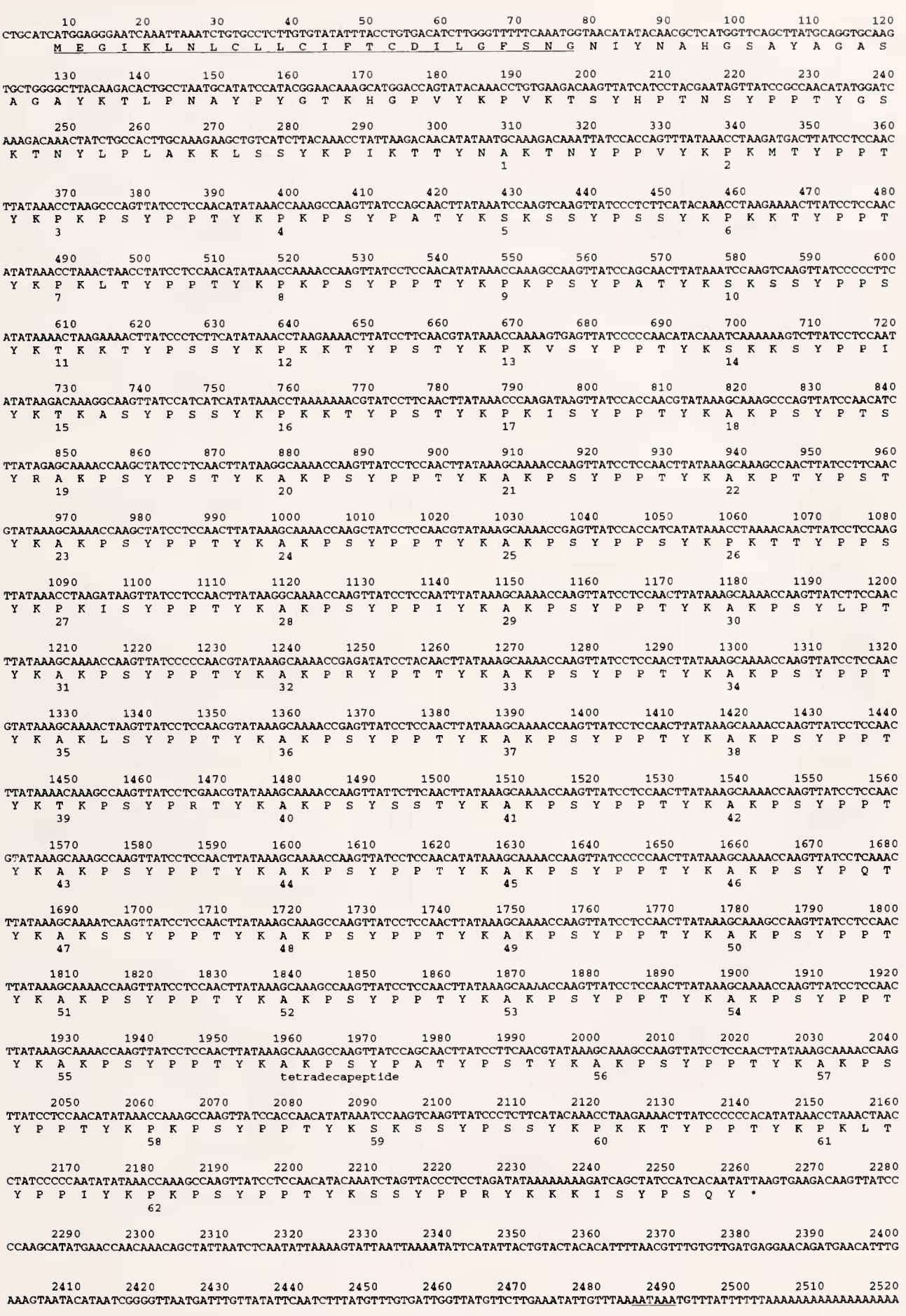
Northern blot hybridization

To examine the efficiency of the probe and to obtain information about the length of the target, northern blot hybridization was carried out. As shown in Figure 1, an intense signal was detected at a position slightly higher than 18S rRNA. This result indicates that the probe is applicable to the screening of the adhesive protein of *M. galloprovincialis*. It also indicates that the target mRNA is expressed in the foot and its length is more than 2.4 kb.

Outline of the structure of the adhesive protein cDNA

About 5×10^4 clones were screened, and more than 50 positive plaques were detected. Of 10 randomly selected clones, 2 were found to have inserts of about 2.5 kb. The longer clone was chosen for further analysis because the shorter one lacked the first several nucleotides (data not shown). Because no restriction site for 16 different enzymes could be found on the insert, deletion derivatives were generated for nucleotide sequence determination. The determined sequence was 2/520 bp, as shown in Figure 2. The coding region determines 751 amino acids, which consist of three distinct parts: the signal peptide of 24 residues, a nonrepetitive region of 76 residues, and a long repetitive region. The amino acid sequence of the signal peptide was similar to that of the *M. edulis* adhesive protein: 22 of 24 residues were conserved between the two species. The amino acid sequence of the nonrepetitive region was also conserved, but several substitutions and a deletion of 18 amino acids were observed (Fig. 3). The repetitive region included 62 repeats of the same decapeptide motif found in *M. edulis*. Although the hexapeptide motif characteristic of *M. edulis* was not observed in the repeats, an irregular tetradecapeptide was seen between the 55th and 56th repeats (Fig. 2, Table I). The sequence of the 3′-untranslated region was also conserved between *M. galloprovincialis* and *M. edulis*, although the termi-

Figure 2. Nucleotide and deduced amino acid sequences of the adhesive protein of *Mytilis galloprovincialis*. Underlined sequence indicates the signal peptide. Also underlined is the polyadenylation signal. Numbers under the amino acid sequence indicate numbers of decapeptide repeats. The asterisks represent the termination codon.


10 20 30 40 50 60 70 80 90 100 110 120
CTGCATCATGGAGGGAATCAAATTAAATCTGTGCCTCTTGTGTATATTTACCTGTGACATCTTGGGTTTTTCAAATGGTAACATATACAACGCTCATGGTTCAGCTTATGCAGGTGCAAG
M E G I K L N L C L L C I F T C D I L G F S N G N I Y N A H G S A Y A G A S
130 140 150 160 170 180 190 200 210 220 230 240
TGCTGGGGCTTACAAGACACTGCCTAATGCATATCCATACGGAACAAAGCATGGACCAGTATACAAACCTGTGAAGACAAGTTATCATCCTACGAATAGTTATCCGCCAACATATGGATC
A G A Y K T L P N A Y P Y G T K H G P V Y K P V K T S Y H P T N S Y P P T Y G S
250 260 270 280 290 300 310 320 330 340 350 360
AAAGACAAACTATCTGCCACTTGCAAAGAAGCTGTCATCTTACAAACCTATTAAGACAACATATAATGCAAAGACAAATTATCCACCAGTTTATAAACCTAAGATGACTTATCCTCCAAC
K T N Y L P L A K K L S S Y K P I K T T Y N A K T N Y P P V Y K P K M T Y P P T
1 2
370 380 390 400 410 420 430 440 450 460 470 480
TTATAAACCTAAGCCCAGTTATCCTCCAACATATAAACCAAAGCCAAGTTATCCAGCAACTTATAAATCCAAGTCAAGTTATCCCTCTTCATACAAACCTAAGAAAACTTATCCTCCAAC
Y K P K P S Y P P T Y K P K P S Y P A T Y K S K S S Y P S S Y K P K K T Y P P T
3 4 5 6
490 500 510 520 530 540 550 560 570 580 590 600
ATATAAACCTAAACTAACCTATCCTCCAACATATAAACCAAAACCAAGTTATCCTCCAACATATAAACCAAAGCCAAGTTATCCAGCAACTTATAAATCCAAGTCAAGTTATCCCCCTTC
Y K P K L T Y P P T Y K P K P S Y P P T Y K P K P S Y P A T Y K S K S S Y P P S
7 8 9 10
610 620 630 640 650 660 670 680 690 700 710 720
ATATAAAACTAAGAAAACTTATCCCTCTTCATATAAACCTAAGAAAACTTATCCTTCAACGTATAAACCAAAAGTGAGTTATCCCCCAACATACAAATCAAAAAAGTCTTATCCTCCAAT
Y K T K K T Y P S S Y K P K K T Y P S T Y K P K V S Y P P T Y K S K K S Y P P I
11 12 13 14
730 740 750 760 770 780 790 800 810 820 830 840
ATATAAGACAAAGGCAAGTTATCCATCATCATATAAACCTAAAAAAACGTATCCTTCAACTTATAAACCCAAGATAAGTTATCCACCAACGTATAAAGCAAAGCCCAGTTATCCAACATC
Y K T K A S Y P S S Y K P K K T Y P S T Y K P K I S Y P P T Y K A K P S Y P T S
15 16 17 18
850 860 870 880 890 900 910 920 930 940 950 960
TTATAGAGCAAAACCAAGCTATCCTTCAACTTATAAGGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAGCCAACTTATCCTTCAAC
Y R A K P S Y P S T Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P T Y P S T
19 20 21 22
970 980 990 1000 1010 1020 1030 1040 1050 1060 1070 1080
GTATAAAGCAAAACCAAGCTATCCTCCAACTTATAAAGCAAAACCAAGCTATCCTCCAACGTATAAAGCAAAACCGAGTTATCCACCATCATATAAACCTAAAACAACTTATCCTCCAAG
Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P S Y K P K T T Y P P S
23 24 25 26
1090 1100 1110 1120 1130 1140 1150 1160 1170 1180 1190 1200
TTATAAACCTAAGATAAGTTATCCTCCAACTTATAAGGCAAAACCAAGTTATCCTCCAATTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCTTCCAAC
Y K P K I S Y P P T Y K A K P S Y P P I Y K A K P S Y P P T Y K A K P S Y L P T
27 28 29 30
1210 1220 1230 1240 1250 1260 1270 1280 1290 1300 1310 1320
TTATAAAGCAAAACCAAGTTATCCCCCAACGTATAAAGCAAAACCGAGATATCCTACAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAAC
Y K A K P S Y P P T Y K A K P R Y P T T Y K A K P S Y P P T Y K A K P S Y P P T
31 32 33 34
1330 1340 1350 1360 1370 1380 1390 1400 1410 1420 1430 1440
GTATAAAGCAAAACTAAGTTATCCTCCAACGTATAAAGCAAAACCGAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAAC
Y K A K L S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T
35 36 37 38
1450 1460 1470 1480 1490 1500 1510 1520 1530 1540 1550 1560
TTATAAAACAAAGCCAAGTTATCCTCGAACGTATAAAGCAAAACCAAGTTATTCTTCAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAAC
Y K T K P S Y P R T Y K A K P S Y S S T Y K A K P S Y P P T Y K A K P S Y P P T
39 40 41 42
1570 1580 1590 1600 1610 1620 1630 1640 1650 1660 1670 1680
GTATAAAGCAAAGCCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAACATATAAAGCAAAACCAAGTTATCCCCCAACTTATAAAGCAAAACCAAGTTATCCTCAAAC
Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P Q T
43 44 45 46
1690 1700 1710 1720 1730 1740 1750 1760 1770 1780 1790 1800
TTATAAAGCAAAATCAAGTTATCCTCCAACTTATAAAGCAAAGCCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAGCCAAGTTATCCTCCAAC
Y K A K S S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T
47 48 49 50
1810 1820 1830 1840 1850 1860 1870 1880 1890 1900 1910 1920
TTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAGCCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAACCAAGTTATCCTCCAAC
Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T Y K A K P S Y P P T
51 52 53 54
1930 1940 1950 1960 1970 1980 1990 2000 2010 2020 2030 2040
TTATAAAGCAAAACCAAGTTATCCTCCAACTTATAAAGCAAAGCCAAGTTATCCAGCAACTTATCCTTCAACGTATAAAGCAAAGCCAAGTTATCCTCCAACTTATAAAGCAAAACCAAG
Y K A K P S Y P P T Y K A K P S Y P A T Y P S T Y K A K P S Y P P T Y K A K P S
55 tetradecapeptide 56 57
2050 2060 2070 2080 2090 2100 2110 2120 2130 2140 2150 2160
TTATCCTCCAACATATAAACCAAAGCCAAGTTATCCACCAACATATAAATCCAAGTCAAGTTATCCCTCTTCATACAAACCTAAGAAAACTTATCCCCCCACATATAAACCTAAACTAAC
Y P P T Y K P K P S Y P P T Y K S K S S Y P S S Y K P K K T Y P P T Y K P K L T
58 59 60 61
2170 2180 2190 2200 2210 2220 2230 2240 2250 2260 2270 2280
CTATCCCCCAATATATAAACCAAAGCCAAGTTATCCTCCAACATACAAATCTAGTTACCCTCCTAGATATAAAAAAAAAGATCAGCTATCCATCACAATATTAAGTGAAGACAAGTTATCC
Y P P I Y K P K P S Y P P T Y K S S Y P P R Y K K K I S Y P S Q Y *
62
2290 2300 2310 2320 2330 2340 2350 2360 2370 2380 2390 2400
CCAAGCATATGAACCAACAAACAGCTATTAATCTCAATATTAAAAGTATTAATTAAAATATTCATATTACTGTACTACACATTTTAACGTTTGTGTTGATGAGGAACAGATGAACATTTG
2410 2420 2430 2440 2450 2460 2470 2480 2490 2500 2510 2520
AAAGTAATACATAATCGGGGTTAATGATTTGTTATATTCAATCTTTATGTTTGTGATTGGTTATGTTCTTGAAATATTGTTTAAAATAAATGTTTATTTTTTAAAAAAAAAAAAAAAAAA

```
                Signal                          Non-repetitive
Mg     1  MEGIKLNLCLLCIFTCDILGFSNG NIYNAHGSAYAGASAGAYKTLPNAYPYGTKHGPVYK
          *************** * ****** ****** * ********** **********  ****
Me     1  MEGIKLNLCLLCIFTFDVLGFSNG NIYNAHVSSYAGASAGAYKKLPNAYPYGTKPEPVYK

Mg    61  PVKTSY-------------------HPTNSYPPTYGSKTNYLPLAKKLSSYKPIKTTYN
          ******                    ** ****************************
Me    61  PVKTSYSAPYKPPTYQPLKKKVDYRPTKSYPPTYGSKTNYLPLAKKLSSYKPIKTTYN

                Repetitive
Mg   101  AKTNYPPVYKPKMTYPPTYKPKPSYPPTYKPKPSYPATYKSKSSYPSSYKPKKTYPPTYK
          ********************************** **     *** *  **  ** *  *******
Me   119  AKTNYPPVYKPKMTYPPTYKPKPSYPPTYKSKP----TYKPKITYPPTYKAKPSYPPTYK
          1          2          3          4          5          6
```

Figure 3. Comparison of the peptide sequences of the signal region, the nonrepetitive region, and the first part of the repetitive region of adhesive protein from *Mytilus galloprovincialis* with those of *M. edulis* described by Laursen (1992). Asterisks indicate the homology between the corresponding sequences. Me and Mg represent sequences of *M. edulis* and *M. galloprovincialis*, respectively.

nator codons were in different positions and several base substitutions were observed (Fig. 4).

Variation of amino acids in the decapeptide motif

As observed in *M. edulis*, some amino acids in the decapeptide motif were sometimes substituted. Substitutions were frequent in the first 17 and last 5 repeats, but they were less common in the middle of the repetitive region. The variation of amino acids in the first three repeats was identical with that of *M. edulis* (Fig. 3) (Filpula *et al.*, 1990; Laursen, 1992), whereas the fourth repeat differed between the two species; *i.e.*, it was a decapeptide in *M. galloprovincialis* but a hexapeptide in *M. edulis*. Figure 5 lists the frequency of substitutions of each amino acid in the decapeptide motif in the whole repetitive region. The most conserved residues were the two tyrosine residues and the lysine at position 2, which were perfectly conserved. The lysine at position 10 and the proline at position 6 were also highly conserved. Other residues suffered considerable variation. The first alanine, the fourth serine, and the eighth threonine were often replaced with proline, threonine, and serine, respectively.

Table 1

Number of decapeptide, hexapeptide, and tetradecapeptide motifs in adhesive proteins of Mytilus galloprovincialis *and* M. edulis

Motif	*M. galloprovincialis*	*M. edulis*[1]	*M. edulis*[2]
Decapeptide	62	71	72
Hexapeptide	0	13	14
Tetradecapeptide	1	0	0

[1] According to Laursen (1992).
[2] According to Filpula *et al.* (1990).

Codon usage in the decapeptide motif

Among the conserved residues, two tyrosine residues and the lysine at position 10 showed highly specific codon usage. All the tyrosine at position 5 and most of the tyrosine at position 9 were coded by TAT (Fig. 5). Most of the lysine residues at position 10 were coded by AAA, but the AAG codon was not as rare at the position of the second lysine (Fig. 5). In other residues, specific codon usage was also observed. For example, the third and sixth proline residues were preferentially coded by CCA and the seventh proline by CCT. In addition, the fourth serine and the eighth threonine were preferentially coded by AGT and ACT, respectively. The first alanine, which was the most substituted residue in the decapeptide motif, was coded only by GCA. Thus, codon usage pattern was highly specific in several residues in the decapeptide motif.

Discussion

The locations of two tyrosine and two lysine and one of the three proline residues in the decapeptide motif were well conserved in the adhesive protein of *M. galloprovincialis*. These residues are also well conserved in the decapeptide repeats of *M. edulis* (Filpula *et al.*, 1990; Laursen, 1992). Tyrosine and lysine residues are also found in the repeat motifs of other mussels (Rzepecki *et al.*, 1991; Laursen, 1992; Waite, 1992) following the paradigm x-Y* -x-x-x-Y*-K, where Y* denotes tyrosine or DOPA. The presence and location of these residues are thought to be critical for the function of adhesive proteins of mussels.

```
Mg 2234 AAAAAGATCAGCTATCCATCACAATATTAAGTGAAGACAAGTTATCCCCAAGCATATGAA
        ********************  **** ***  *************** ******* **
Me      AAAAAGATCAGCTATCCATCATCATATAAAGCTAAGACAAGTTATCCCCCAGCATATAAA

Mg 2294 CCAACAAACAGCTATTAATCTCAATATTAAAAGTATTAATTAAAATATTCATATTACTGT
        *********** ************************** *********** ********
Me      CCAACAAACAGATATTAATCTCAATATTAAAAGTATTAACTAAAATATTCACATTACTGT

Mg 2354 ACTACACATTTTAACGTTTGTGTTGATGAGGAACAGATGAACATTTGAAAGTAATACATA
        ********************* ****************************************
Me      ACTACACATTTTAACGTTTGTATTGATGAGGAACAGATGAACATTTGAAAGTAATACATA

Mg 2414 ATCGGGGTTAATGATTTGTTATATTCAATCTT--TATGTTTGTGATTGGTTATGTTCTTG
        ********************************  ************ ************
Me      ATCGGGGTTAATGATTTGTTATATTCAATCTTAATATGTTTGTGATTTGTTATGTTCTTG

Mg 2474 AAATATTGTTTAAAATAAATGTTTATTTTTT(Poly A)
        ** ******** ******* ******* ***
Me      AAGTATTGTTTCAAATAAAAGTTTATTCTTTTCTGGT(Poly A)
```

Figure 4. Comparison of the nucleotide sequence from the last part of the repetitive region to the poly(A) tail of *Mytilis galloprovincialis* with the same region of *M. edulis* reported by Strausberg *et al.* (1989). Asterisks indicate the homology between the corresponding sequences. The underlined sequence indicates the stop codon. Me and Mg represent sequences of *M. edulis* and *M. galloprovincialis*, respectively.

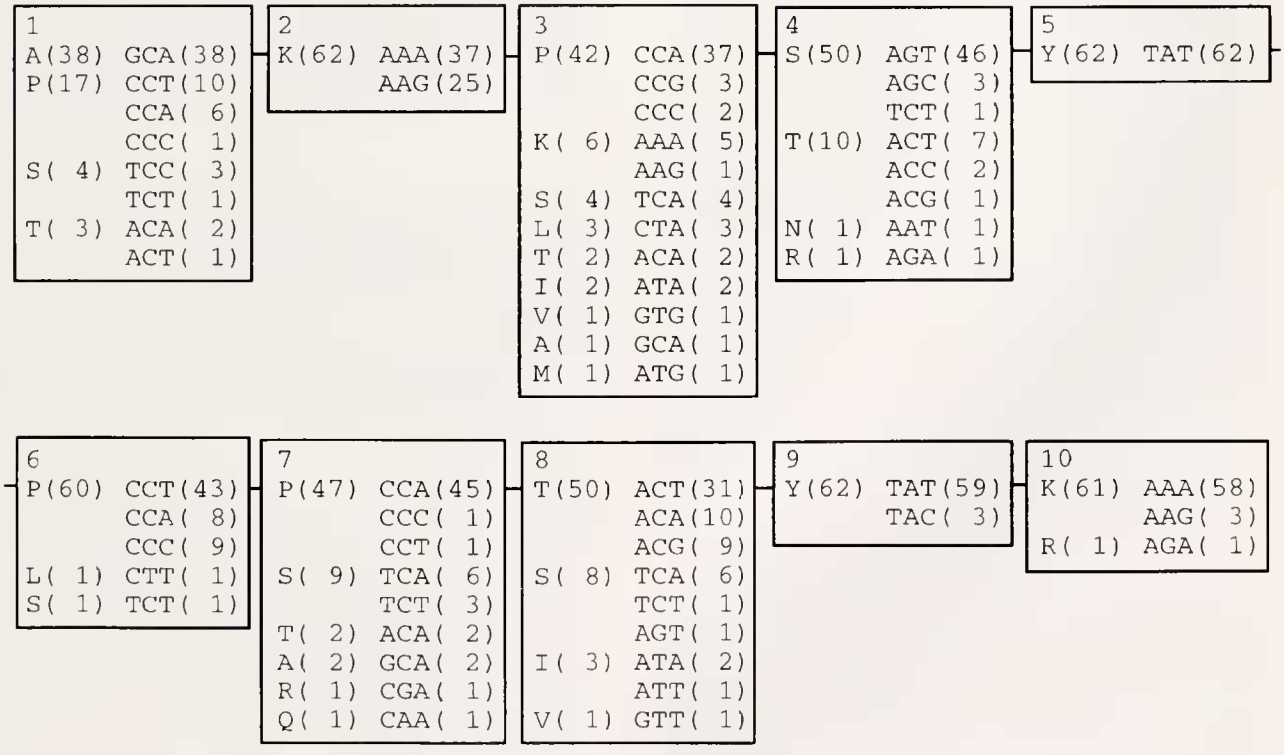


Figure 5. Frequencies of amino acid substitutions and codon usages in the decapeptide motif of the adhesive protein of *Mytilis galloprovincialis*. Numbers in parentheses indicate the frequencies of each of the amino acids and codons, respectively.

In the previous and present studies, three motifs—decapeptide, hexapeptide, and tetradecapeptide—were observed. We noticed that these motifs can be divided into two submotifs, (Y)KAKPSY (submotif A) and (Y)PPTY (submotif B), according to the position of the tyrosine residues. The hexapeptide, decapeptide, and tetradecapeptide motifs corresponded to A, A + B, and A + B + B, respectively, though minor variations were observed in the positions of Ala, Pro, Ser, and Thr residues. The decapeptide motif is obviously the basic motif of the adhesive protein. The hexapeptide motif is apparently also a functional unit, or at least it does not prevent the function, because it is not rare in the repetitive region of *M. edulis*. The tetradecapeptide may also be a functional unit because it is composed of the same submotifs.

In the nucleotide sequence of the repetitive region, specific codon usage was observed. It is interesting that the codon usage in one of the two lysine residues was highly specific but the other was not. In addition, the alanine residues were often replaced with proline or other amino acids, but all the alanine residues at this position were coded only by GCA. The third proline was also frequently substituted by various amino acids, but the CCA codon was preferentially used for proline, and the codons whose third bases are A were preferentially used for other amino acids. The same tendencies were observed in the gene for the adhesive protein of *M. edulis* (Filpila *et al.*, 1990). These conserved nucleotides suggest that the specific nucleotide sequences have some functional significance (*e.g.*, in the transcriptional regulation as reported in fibroin mRNA (Mita *et al.*, 1988) or in the replication of genome); but information is insufficient for discussion at present.

M. galloprovincialis is thought to have originated in the Mediterranean Sea and to have been accidentally introduced into Japan (Wilkins *et al.*, 1983). Because it has many morphological and genetical characteristics in common with *M. edulis*, these two species have been thought to be closely related (Seed, 1992, for a review) or even to be subspecies of *M. edulis* (Gosling, 1984; Gardner, 1992; for reviews). Even with the use of mitochondrial ribosomal DNA sequences (Geller *et al.*, 1993), it is difficult to distinguish between these two species. They appear to maintain genetic differentiation, however, even though hybridization is possible between them (McDonald *et al.*, 1991). Two different sequences of the adhesive protein from two different strains have been reported; one is derived from the cDNA sequence described by Laursen (1992), and the other is from the partial genomic sequence by Filpula *et al.* (1990). The latter lacks the N-terminal sequence, but the former sequence from the 53th residue to the end of the nonrepetitive region was identical with the corresponding sequence of the latter at the amino acid level. The signal region and the nonrepetitive region of *M. galloprovincialis* had amino acid sequences similar to but not identical with those of *M. edulis*. It has been reported that two *M. edulis* sequences in the repetitive region were identical in the first nine and last five repeats and only the distribution pattern of hexapeptides in the middle of the repetitive region was different. The hexapeptides that exist in the repetitive region of *M. edulis* were not found in the sequence of *M. galloprovincialis;* a tetradecapeptide was found instead. We determined partial sequences of other cDNA clones of *M. galloprovincialis*, but also failed to find any hexapeptides (data not shown). However, more information is required before we can discuss the correlation of sequence differences with diversity among populations and species. We are now using polymerase chain reaction to look for interspecific and intrapopulation variation in adhesive protein sequences. The sequence of the adhesive protein may offer a key to understanding not only the function of this protein but also the genetic diversity among different populations and species of mussels.

Acknowledgments

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